Lord Howe Island Psocoptera (Insecta)

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A brief history of collecting Psocoptera on Lord Howe Island is given. One new genus, Mauropsocus (Pseudocaeciliidae: Zelandopsocinae), and three new species, Mauropsocus monteithi, Lepolepis trifasciata (Lepidopsocidae) and Nimbopsocus huttoni (Myopsocidae) are described. An additional new genus and species is noted but not formally described and named because adult material has not yet been found. Twentyseven species of Psocoptera are now known from Lord Howe of which fourteen are probably endemic to the island. Of the thirteen non-endemic species eight have widespread distributions beyond the island (some being found in domestic situations); three occur otherwise only in Australia and one is known only from Norfolk Island. One species occurs in Australia, New Caledonia, Norfolk Island and New Zealand. It is anticipated that the fauna is not yet completely known but most of the endemic species are members of the related families Pseudocaeciliidae, Philotarsidae and Elipsocidae. Distribution of the species is summarized in a table.


The first recorded psocopteran from Lord Howe Island, a female specimen of Aaroniella howensis Smithers & Thornton, was collected by Ms Zenta Liepa in 1955 and described in 1975. This and material collected by the author in November 1969, February 1971 and August 1971 and by Geoffrey Holloway in September 1971 and March 1974 formed the basis of the first paper on the Psocoptera of Lord Howe Island (Smithers & Thornton, 1975). The author visited the island again in February 1977 (Smithers, 1979). Between September 1978 and October 1979 Tim Kingston collected Psocoptera as part of a general insect survey and in 1979 Geoff Monteith, of the Queensland Museum, made what was then the most comprehensive collection of Psocoptera from Lord Howe Island (Smithers & Thornton, 1975). Kingston made a small collection in 1980 and Patrick Huber, in 1992–1993, collected a few specimens during a survey of insects associated with the inflorescences of the economically important Thatch Palm, Howea forsteriana (C. Moore and F. Muell.) Becc. (Smithers, 1995). The biggest collection to date is that made during the Lord Howe Island Invertebrate Biodiversity Survey in which several collectors, based at the Australian Museum, using a variety of collecting techniques, amassed a substantial amount of material during several visits. Since then Ian Hutton has provided additional material collected between 2000 and 2003. Some of the material from the Lord Howe Invertebrate Biodiversity Survey has no designated individual collector; in the lists of material studied in this paper these specimens are referred to as being collected by LHIS.

Except where indicated otherwise specimens collected by Monteith will be returned to the Queensland Museum. Other material is in the Australian Museum. Nymphs are not designated as type material.

Full synonymies and references to the species can be found in Smithers (1967, 1996) and Lienhard and Smithers (2002) or other references given in this paper. Plant names used are as in Wilson (1994), with later amendments where necessary. Vegetation types mentioned in association with some of the material collected by Monteith are described by Pickard (1983); these are referred to as Pickard veg on the specimen labels and hence in the text of this paper.

In the species descriptions the following abbreviations are used: F = length of hind femur; T = length of hind tibia; t1,
Comparison with collecting experience on Norfolk Island suggests that it is very likely that there are more species to be recorded from Lord Howe Island. Norfolk Island has about twice the land area of Lord Howe Island (about 34 square kilometres compared to 16 square kilometres for Lord Howe). Although Lord Howe and Norfolk have about the same number of vascular plants (459 and 445 species respectively) Lord Howe has a larger indigenous flora than Norfolk (241 spp. of vascular plants as opposed to 171 spp.) (Wilson, 1994), a much more varied topography (875 metres altitude as opposed to 316 m.) and although subjected to less collecting effort by specialist collectors it has yielded a comparable number of species of Psocoptera (27, of which 14 appear to be endemic) to that obtained on Norfolk (21, of which 11 appear to be endemic). Fifteen species of Psocoptera have been added to the Lord Howe list in this paper whereas only five additional species have been found on Norfolk since 1978, the latest addition (in 1998) being a single, widely distributed tropical species (Smithers, Peters & Thornton, 2000). Despite its greater (but much more disturbed) area it seems likely that most species have now been recorded for Norfolk. On the other hand, circumstances on Lord Howe suggest that more species will be found there. This possibility should be remembered when considering relationships of the Lord Howe fauna to those of other areas (Table 1).

Table 1. Distribution of Lord Howe Island Psocoptera.

<table>
<thead>
<tr>
<th>Lord Howe species</th>
<th>endemic&lt;sup&gt;1&lt;/sup&gt;</th>
<th>widespread&lt;sup&gt;4&lt;/sup&gt;</th>
<th>Australia</th>
<th>Norfolk Island</th>
<th>New Caledonia</th>
<th>New Zealand</th>
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<td>Lepidopsocidae</td>
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<td>Lepolepis trifasciata&lt;sup&gt;1&lt;/sup&gt;</td>
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<td>Paracaecriulus lemurisi&lt;sup&gt;1&lt;/sup&gt;</td>
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<td>Nimbopsocus australis&lt;sup&gt;1&lt;/sup&gt;</td>
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<td>number of species: 27</td>
<td>14</td>
<td>9</td>
<td>11</td>
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In the table: • = species is present in the area indicated at the head of the column;
<sup>1</sup> first record for Lord Howe in this paper;
<sup>2</sup> species frequently associated with human domestic environment;
<sup>3</sup> known so far only from Lord Howe Island;
<sup>4</sup> occurs in areas other than above, in some cases almost cosmopolitan, see text for details.
Systematic treatment of Lord Howe Island Psocoptera

**LEPIDOPSOCIDAE**

*Echmepteryx anomala* Smithers & Thornton


**Distribution.** Known only from Lord Howe Island.

*Echmepteryx (Loxapholia) howensis* Smithers & Thornton


**Echmepteryx (Thylacopsis) madagascariensis** (Kolbe)


**Distribution.** First record for Lord Howe Island. Previously known from Madagascar, Réunion, Isle Glorieuse, Seychelles, East Africa, Ivory Coast, Bioko Is., South America, North America, Australia, Hawai, Bonin Is., Marianas, Kermadecs, Chagos Archipelago, New Zealand, Norfolk Island, Galapagos, Tonga, Society Islands, Indonesia, Bermuda, Jâmaica, Fiji, Diego Garcia, Chile, Hong Kong, Germany (introduced, in greenhouse).

**Lepolepis trifasciata n.sp.**

Mt Gower summit on ridge between creeks 3 (most western) and 2 (middle), Pittosporum erioluma, Zygogynum, Metrosideros nervulosa, leaf litter, 25.vi.2001, Ian Hutton. 1♀, Lago Beach between rubbish tip and airstrip, closed rain forest, Drypetes/Cryptocarya, pitfall trap, 27.xi.–4.xii.2000, LHIIS. 1♀, western slopes of Roach Island, orthophyll short grass, Poo, pitfall trap, 29.xi.–7.xii.2000, LHIIS. 1♀, Transit Hill, 60 m, leaf litter, Chionanthus quadristaminus, Cleistocalyx fallagaraii, 26.x.1979, T. Kingston. 2♀, same data, 10.x.1978, T. Kingston. 1♀, same data, 26.x.1979, T. Kingston. 1♀, same data, 8.iv.1979, T. Kingston. 1♂, 1♀, Clear place, leaf litter, 6.i.1980, T. Kingston. 1♂, south end, Settlement Beach, under bark, Lagunaria, 7.viii.2000. No collector. 1♀, 2m, Transit Hill, north slope, Queensland Museum berlesate 151, volcanic soil, Pickard veg DaCt, 18.xi.1979, G.B. Monteith. 1♂, 1♀, 2m, same data, Queensland Museum berlesate 152, volcanic soil, Pickard veg DaCt, sieved litter, 18.xi.1979, G.B. Monteith. 1♂, same locality, west base, Queensland Museum berlesate 118, volcanic soil, 5 m, Pickard veg DaCt, sieved litter, 4.xi.1979, G.B. Monteith. 1♂, same locality, north slope, Queensland Museum berlesate 137, Pickard veg CfLq, sieved litter, 18.xi.1979, G.B. Monteith. 1♂, 3♀, North Bay, 5 m, leaf litter, Howea forsteriana, 15.xi.1978, T. Kingston. 1♀, North Bay, west end, 5 m, Queensland Museum berlesate 156, Pickard veg Hf, sieved litter, volcanic soil, 19.xi.1979, G.B. Monteith. 3♂, 2♀, Old Settlement, 80 m, litter, Drypetes deplanchei, Cryptocarya triplinervis, 18.iv.1979, T. Kingston. 3♂, 5♀, same data, 2.vi.1979, T. Kingston. 1♂, same data, 8.iv.1979, T. Kingston. 1♂, 1♀, Stevens Reserve, 5 m, leaf litter, Howea forsteriana, 1.x.1978, T. Kingston. 1♂, 1♀, same data, LHI 168, No 8, 25.ix.1978, T. Kingston. 1♂, same data, LHI 191, No 8, T. Kingston. 1♂, 2♀, same data, 30.ix.1978, T. Kingston. 1♂, Intermediate Hill, leaf litter, Cleistocalyx fallagaraii, Chionanthus quadristaminus, LHI 327, No 8, 24.i.1979, T. Kingston. 2♀, same locality, LHI 316, No 8, 19.1.1979, T. Kingston. 5♂, 1♀, same locality, North Hummock, Queensland Museum berlesate 125, volcanic soil, 180 m, Pickard veg CfLq, sieved litter, 6.xi.1979, G.B. Monteith. 1♂, 1♀, Intermediate Hill, Cleistocalyx fallagaraii, Chionanthus quadristamineus, 100 m, LHI 347, No 8, 24.i.1979, T. Kingston. 9♂, 11♀, 31nn, S of Clear Place, 6.ii.1980, T. Kingston. 1♂, Mt. Gower, leaf litter, 100 m, no date, T. Kingston. 1♀, Mt. Gower, 350 m, ii.1979, T. Kingston. 1♂, 1♀, Lord Howe Island Stations: 007–0037, no date, T. Kingston. 1♂, Erskine Valley, leaf litter, no date, T. Kingston. 1♀, Erskine Valley, 175 m, leaf litter, Drypetes deplanchei, Cryptocarya triplinervis, LHI 454, No 8, 1.viii. T. Kingston. 4♂, 1♀, 10m, Broken Banyan Apartments, leaf litter, 7.x.1994, D.S. Horning and D. Horning. 3♀, Lord Howe Island, no date, D. Horning. 1♂, Smoking Tree Ridge summit, Queensland Museum berlesate 130, volcanic soil, 150 m, Pickard veg DaCt, sieved litter, 7.xi.1979, G.B. Monteith. 1♂, same locality, east face, Queensland Museum berlesate, 163, volcanic soil, Pickard veg CfLq, sieved litter, 23.xi.1979, G.B. Monteith. 2♂, 6♀, Gully at Catalina crash, Queensland Museum berlesate 169, volcanic soil, 20 m, Pickard veg Hb, sieved litter, 25.xi.1979, G.B. Monteith. 1♂, 1♀, Lagoon Road, opposite museum, Queensland Museum berlesate 122, alluvial soil, Pickard veg DaCt, sieved litter, 5.xi.1979, G.B. Monteith. 5♂, 10♀, Lagoon Road, opposite hospital, Queensland Museum berlesate 119, alluvial soil, 2 m, Pickard veg DaCt, sieved litter, 4.xi.1979, G.B. Monteith. 1♂, 1♀, Malabar summit, Queensland Museum berlesate, 168, volcanic soil, 200 m, Pickard veg DaCt, sieved litter, 25.xi.1979, G.B. Monteith. 1♀, Boat Harbour, Queensland Museum berlesate 161, 10 m, volcanic soil, Pickard veg CfLq, sieved litter, 23.xi.1979, G.B. Monteith. 1♂, Mount Eliza summit, Queensland Museum berlesate 154, volcanic soil, 150 m, Pickard veg MnCa, sieved litter, 19.xi.1979, G.B. Monteith. 1♂, Dawsons Point Ridge summit, 150 m, Queensland Museum berlesate 120, Pickard veg DaCt, sieved litter, 5.xi.1979, G.B. Monteith. 1♂, Little Slope, 50 m, leaf litter, Howea forsteriana, LHI 355 No. 8, 22.xi.1979. T. Kingston. 4♂, 5♀, Behind Leanda Lei, Queensland Museum berlesate 127, calcareous soil, Pickard veg Hf, sieved litter, 6.xi.1979, G.B. Monteith.

Specimens collected by other than G.B.Monteith are in the Australian Museum; specimens collected by G.B.Monteith are deposited in the Queensland Museum.
Description

Female. Coloration (in alcohol). This species exhibits extreme variation in depth of colour, ranging from pale testaceous to specimens which are almost black. The variation may be age-related; nymphs are very pale. Parts are consistently relatively darker than one another through the series of specimens. Vertex, postclypeus, labrum and maxillary palps a little darker than the frons, genae darkest. Antennae very pale to dark brown. Eyes black. Thoracic nota variable, pleura darker. Legs variable with tarsi always paler than other segments. Fore wings (Fig. 1) pale with basal, middle and preapical irregular brown bands, the colour varies in depth to similar degree as other parts of the body of the same specimen. Abdomen dorsally pale in basal half, where it is covered by the reduced wings, distally darker where it is exposed. Terminal structures dark.

Morphology. Brachypterous. Length of body: 2.0 mm. Median epicranial suture distinct, anterior arms very short, evanescent near origin. Vertex sharp. Postclypeus very slightly bulging. Head strongly pubescent, genal setae especially long and stout. Top of eyes about level with vertex. IO/D: 2.3; PO: 0.8. Ocelli absent. Labrum (Fig. 3) with five distal inner labral sensilla. Lacinia (Fig. 7). Measurements of hind leg: F: 0.62 mm; T: 0.73 mm; t1: 0.29 mm; t2: 0.07 mm; t3: 0.06 mm; rt: 4.1:1:0.85. No ctenidiobothria. Fore wing length: 1.2 mm; width: 0.5 mm. Fore wing (Fig. 1) somewhat elytriform, veinless. Costal and hind margins with thickened band in basal half of wing. Wing surface with vestiture of very narrow scale-like setae as well as fairly dense cover of well-developed, erect, setae (nearly all lost in preservative). Wings reach to about three quarters of length of the abdomen. Hind wings reduced to a small, membranous, flap. Epiproct (Fig. 2) simple, rounded behind, sparsely setose. Paraproct simple, two trichobothria with ornamented alveoli and a few setae in distal half. Posterior spine slightly downwardly curved. Subgenital plate simple, wider than long, sparsely setose. Gonapophyses (Fig. 4). Spermathecal sac membranous, apparently without dentate sclerotic ring and without maculae. Associated with the sac is a large, strongly sclerotized, partly hollow, peg-like structure (Fig. 5, Fig. 6 [enlarged]). The structure is pointed at one end, widened at the other and hollow almost to the pointed end. The wider end of the funnel thus formed is open along one side. The peg-like structure appears to arise in and be an integral part of the otherwise thin, membranous wall of the sac. It lies in an outwardly-directed tube-like extension of the wall the mouth of which is firmly attached to the peg about half way along it so that part of the peg appears to be inside and part outside the sac tube. There are folds at the base of the tube which suggest that the peg is capable of being moved inwards and outwards along the lumen of the tube whilst being attached to its inner wall. The structure is seen in various positions through the abdominal wall, depending on the position occupied by the spermathecal sac and is possibly equivalent to the sheath of the spermathecal duct opening of Mockford (Mockford, 2005). It is large enough to be seen as a sclerotized, peg-like rod in the undissected abdomen.


Morphology. General morphology as in female. Length of body 2.2 mm. IO/D: 2.4; PO: 0.86. Fore wing length: 0.98 mm; width: 0.46 mm. Measurements of hind leg: F: 0.60 mm; T: 0.68 mm; t1: 0.29 mm; t2: 0.07 mm; t3: 0.06; rt: 4.1:1:0.85. Ctenidiobothria absent. Epiproct as in female, simple, rounded behind, sparsely setose. Paraproct simple, two trichobothria with ornamented alveoli and a few setae in distal half. Hypandrium simple, rounded behind, setose. Phallosome (Fig. 8).

Discussion

There are now seven species described in the genus Lepolepis Enderlein: L. bicolor Broadhead (England (on introduced ground nuts from Africa), Ile Glorieuse, Réunion, India), L. ceylonica Enderlein (Sri Lanka, Taiwan), L. columbiensis Badonnel (Colombia), L. graemei Smithers (Norfolk Is.), L. pateriformis New (Alzabra), L. picta Thornton (Hawaii), and L. trifasciata (Lord Howe Island). The wings of L. graemei and L. trifasciata are longer than in the other species. When L. graemei was described no mention was made of the conspicuous peg-like structure associated with the spermatheca and reexamination of the dissection of the genitalia of a female paratype did not reveal any such organ. Having found the organ in the obviously similar L. trifasciata I reexamined further material of L. graemei and found that an almost identical structure was, in fact, present. The spermathecal sac in this genus is very delicate, was probably damaged and the peg-like structure lost when the earlier paratype was dissected. The peg-like structure is not easily homologized with part of the reproductive organs of any other female psocopterans but, as mentioned above, there is a possibility of it being homologous to the sheath of the spermathecal duct opening of Mockford (Mockford, 2005). The spermatheca has not been described for all species of Lepolepis but L. graemei and L. trifasciata share several significant features with each other which they do not share with other members of the genus for which the spermatheca has been described, such as a lesser degree of wing reduction, similarities in wing pattern, the presence of the peg-like structure associated with the spermatheca and the lack of spermathecal maculae, which are present in some of the other species. This suggests that the two species are closely related and stand apart from the others of which the spermatheca has been described. Given that the presence of the peg-like structure is very likely a shared apomorphy it may be reasonable to erect a new genus for their accommodation.

I hesitate to do this until further information is available on the nature of the spermatheca in those species for which it has not yet been described.

Etymology. The specific name refers to the three broad transverse dark bands across the wings.

Distribution. Known only from Lord Howe Island.
TROGIIDAE

**Cerobasis annulata** (Hagen)


**Material studied.** 1 ♀, Mount Gower, litter, 850 m, mossy forest, 27.ix.1978, T. Kingston. 1 ♀, Erskine Valley, 175 m, litter, *Drypetes, Cryptocarya*. 12.viii.1979, T. Kingston.

**Distribution.** First record for Lord Howe Island. Previously known from Europe, North America, Australia (in stored products), St Helena, Robinson Crusoe Island, Hawaii, Azores, Morocco, Madeira. In the wild and in domestic situations.

**Cerobasis guestfalicus** (Kolbe)


**Pseudoneuropt Russ. Reich** p. 496.


**Distribution.** First record for Lord Howe Island. Previously known from North America, Europe, Canary Is., Azores, Israel, Saudi Arabia, Sardinia, Morocco, Tunisia, Japan, Mexico, Java, Australia, Norfolk Is., New Zealand, Hawaii, Bermuda, Kenya, South Africa, Robinson Crusoe Is., Argentina, Brazil, Chile, St Helena, St Paul Is., Jamaica, Kermadecs, Mauritius. In the wild and in domestic situations.

**Lepinotus inquilinus v. Heyden**

Clothilla inquilina (Heyden). Brauer & Löw, 1857.

**Synonymy:** Enderlein, 1905. *Res. Swedish Exp. Egypt and White Nile* p. 25.


**Distribution.** First record for Lord Howe Island. The large number of previously published records indicate an almost cosmopolitan distribution for this species which is found indoors and in the wild.

**PSOQUILLIDAE**

**Rhyopsocidus Niger** (Smithers)


**Distribution.** Recorded only from Lord Howe Island (as *Trogium nigrum* Smithers).
**PSYLLIPSOCIDAE**

*Psocathropos lachlani* Ribaga


**Distribution.** First record for Lord Howe Island. Also known from Australia, Europe, Morocco, North America, Cuba, Jamaica, Nicaragua, Mozambique, Angola, Congo, Nigeria, Madeira, Madagascar, Réunion, Thailand, Taiwan, Hawaii. Found in the wild and in domestic situations.

**CAECILIUSIDAE**

*Stenocaecilius quercus* (Edwards)


**Distribution.** Previously recorded from Lord Howe Island, Tasmania and Australia.
Paracaecilius lemuris Smithers


Distribution. First record for Lord Howe Island. Previously recorded from Australia.

Valenzuela pteridii (Smithers)


Material studied. 1♀, Rocky Run Creek, where Intermediate Hill track crosses, litter, Cleistocalyx fullagarii, Pandanus forsteri, Coprosma huttoniana, 18.v.2002, I. Hutton.

Distribution. First record for Lord Howe Island. Previously recorded from the Australia and Tasmania (as Caecilius pteridii).

ECTOPSISOCIDAE

Ectopsocus petersi Smithers


Distribution. Previously recorded from Lord Howe Island (as Ectopsocus punctatus Thornton and Wong). Also recorded from Great Britain, Europe, North America (introduced?), Egypt, Australia, New Zealand. Common on dead leaves.

Ectopsocus insularis Smithers & Thornton


Distribution. First record for Lord Howe Island. Previously recorded from Norfolk Island.
**PERIPSOCIDAE**

*Peripsocus milleri* (Tillyard)


**Distribution.** First record from Lord Howe Island. Previously recorded from Tasmania, Australia, New Zealand, Auckland Island, Kermadecs, Norfolk Island, Hawaii, Europe, Canary Islands, Madeira, North America. It is also known to occur in New Caledonia (unpublished material in the Australian Museum). Corticulous.

*Peripsocus similis* Enderlein


**Distribution.** Known only from Lord Howe Island.

*Mepleres fasciatus* (Smithers & Thornton)


**Distribution.** Known only from Lord Howe Island.

*Mepleres hollowayi* (Smithers & Thornton)


**Mauropsocus n.gen.**

Belonging to the Pseudocaeciliidae: Zelandopsocinae. Females micropterous. Tarsi 3-segmented. Claws with a minute denticle on one claw of each pair. Subgenital plate incipiently bilobed, each lobe with one large posteriorly directed seta near hind margin. Ventral valve and dorsal valve of gonapophyses with large, pointed apophysis. Setae on head, thorax and abdomen (other than genitalia) include many seated in large, raised alveoli and have distal ends truncate, expanded or extended on one side at the apex. No ocelli. Apical antennal segment distally narrow. Abdominal terga each with well sclerotized posterior transverse band and a more lightly sclerotized anterior band.

**Type species:** *Mauropsocus monteithi* n.sp.

*Mauropsocus* differs significantly from other genera of the subfamily in having many of the setae conspicuously modified. They arise from greatly exaggerated, raised alveoli and many have the apex of unusual form. The apex of some setae is simply truncate, in others it is distally expanded or the apex is modified so that one side of the apex is extended beyond the other (Figs 10–14). The modified setae are also much thicker than normal setae and in many the base is also modified, being narrowed, neck-like, at the attachment to the alveolus (Fig. 13). The male of *M. monteithi* is not known but on the basis of the female characters alone its nearest relatives appear to be *Zelandopsocus*, *Austropsocus* and *Howeanaum*. *Howeanaum* is known from two species, *H. costale* (Thornton and New) (Australian) and *H. huberi* Smithers (Lord Howe Island). The former is macropterous in both sexes and the latter is macropterous in the male and micropterous in the female.

**Mauropsocus monteithi** n.sp.

**Material studied.** 1 ♀ (HOLOTYPE), QM T99350, Big Creek, Mountain Inn, rainforest, pyrethrum knockdown, 11.xi.1979, G.B. Monteith. Holotype in the Queensland Museum.

**Etymology.** Mauros = Gr. dark, referring to the dark colour of the species. This species is named for Dr Monteith in recognition of his immense contribution to Australian entomology.

**Distribution.** Known only from Lord Howe Island.

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**Description**

**Female.** Coloration (in alcohol). Head dark brown, a paler spot in middle of front of head, in position usually occupied by ocelli. A darker mark from eye to antenna base, lateral and anterior transverse margin of postclypeus very dark brown. Median epicranial suture almost black; anterior arms absent. Postclypeus with indistinct, irregular, almost transverse, slightly converging dark stripes. Labrum pale. Antennae brown. Eyes black. Maxillary palps brown. Prothorax dorsally dark. Meso- and metanotum each with
dark brown transverse band across posterior part of tergum in which dark spots are arranged in irregular transverse rows, the spots being large alveoli from which arise strong setae. Anterior part of tergum paler. Legs dark brown except for distal half of hind tibia and tarsus, which are slightly paler. Abdominal terga banded in similar way to thoracic terga i.e each tergum darker in posterior part in than in anterior part. A fine, pale, median longitudinal line from front of mesonotum to hindmost abdominal tergum.

**Morphology.** Micropterous. Length of body: 1.7 mm. Median epicranial suture very distinct, finer anteriorly, anterior arms absent. Head capsule with raised, enlarged alveoli giving most of head a rough appearance. Vertex narrow but rounded. Front of head flat. Frons poorly delimited because of lack of anterior arms of epicranial suture and posterior part of epistomial suture. Head from side (Fig. 9, setae not shown) short, postclypeus hardly protruding. Labrum without lateral styli on anterior margin. Five inner sensilla, two trichoid, three placoid. Genae glabrous, without enlarged alveoli, therefore having a smoother appearance than rest of head capsule. Antennae 13-segmented but short; apical segment strongly narrowed distally. Length of flagellar segments: f1: 0.055 mm.; f2: 0.045 mm. Eyes very small, round, of about 20 spaced ommatidia, placed on side of head slightly nearer to base of antenna than vertex. No ocelli. Fourth segment of maxillary palp short, width two-thirds of length. Thoracic and abdominal terga each with sclerotized transverse band in posterior part giving the dorsal view of the insect its strongly banded appearance. Integument of each segment anterior to the sclerotized band less heavily sclerotized. More heavily sclerotized part of the terga bears rows of strongly developed, variously modified setae (Figs 10–12) each arising from an enlarged alveolus. Mesonotum with one posterior row of setae and two single setae representing an anterior row. Integument in heavily sclerotized posterior areas bears more or less evenly spaced tiny papillae. Abdominal terga as thoracic but surface papillae arranged roughly in transverse rows in some areas. Modified setae in two irregular rows, those of the posterior row on each segment larger than those of anterior row. Measurement of hind leg: F: 0.26 mm.; T: 0.39 mm.; t1: 0.07 mm.; t2: 0.02 mm.; t3: 0.04 mm.; r: 3.5:1.2. No ctenidiobothria. Claws with one minute denticle on one claw of each pair. Outer side of femora sparsely setose, setae fine. Fore tibia bearing fine, normal setae overall, outer side with a row of a few larger, blunt-ended setae arising from larger raised alveoli. Mesotibia as front tibia but the row of setae on the outer side has expanded truncate tips. Metatibia as mesotibia but larger setae with expanded truncate apices in a row on the outer side arise from large alveoli. Pulvillus broad. No coxal organ. Fore wing (Fig. 14) reduced to a very small lobe with a single, very large, modified apical seta (Figs 13, 14). Hind wing (Fig. 15) smaller, simple, without seta. Epiproct (Fig. 16) a simple, lightly sclerotized lobe with a transverse ridge bearing a few long, fine setae. Paraproct (Fig. 17) lightly sclerotized, with a few setae. Subgenital plate (damaged in preparation) apparently incipiently bilobed behind with a large seta on each lobe. Gonapophyses (Fig. 18). Ventral and dorsal valves each with well-developed, long, fine, preapical apophysis.

**Male.** Unknown.

**Discussion**

Mockford (1984) reassessed and characterized the genera then included in the Philotarsiidae and some of the genera in the Pseudocaeciliidae. As a result he made some redistribution of the genera between the families and erected the family Bryopsocidae (for *Bryopsocus* Thornton, Wong and Smithers) which he considered to be intermediate between the other two families.

*Mauropsocus monteithi* is clearly a micropterous pseudocaeciliid belonging to the subfamily Zelandopsocinae and it is appropriate to compare it with the five genera of the family which have 3-segmented tarsi, namely, *Zelandopsocus* Tillyard, *Austropsocus* Smithers, *Novopsocus* Thornton, *Howeanum* Smithers and *Trimerocaecilius* Meinander, although the last is not considered a member of the Zelandopsocinae and may not even be a true pseudocaeciliid (Lienhard, 1998).

**Pseudocaeciliidae n.gen. and n.sp.**

The collection includes two remarkable nymphs which cannot readily be associated with any adults so far collected from Lord Howe Island. They certainly represent an undescribed genus and species. The most likely family to which they belong is the Pseudocaeciliidae. As adults are not yet available this placing must be considered tentative.

**Material studied.** *In*, Southern end of Salmon Beach, vic. Little Island, *Xylosoma maidenii*, beating, 27.xi.2000, C. Reid, H. Smith. *In*, Stephens Reserve, New Settlement, *Drypetes deplanchei*, beating, 13.xii.2000, R. Harris. (Originally preserved on points these two specimens have been removed and are now stored in alcohol).

**Description**

**Nymph.** The following brief description is based mainly on the larger specimen.

**Coloration.** Head pale straw-coloured, with slightly darker, irregular, band adjacent to the inner margin of the compound eyes; a similar band on either side of the median
epicranial suture from vertex to position usually occupied by the ocellar tubercle. Postclypeus, genae and labrum straw-coloured. Eyes colourless. Antennae coloured as head. Mesonotum mottled, irregularly straw-coloured; metanotum pale with a brownish spot on each side of midline. Legs colourless except for black claws. Abdominal tergites 4–6 brownish, other tergites pale.

Morphology. Antennal flagellar segments with well-developed, porect setae, many longer than segment diameter. Head almost circular in frontal view; front of head, including frons and postclypeus, strongly concave so that the front of the head appears hollowed out or bowl-shaped. When viewed from the side the head is very short, the anterior and posterior margins almost parallel to one another, the head as a whole thus somewhat discoid. Epistomial suture not obvious. Rim of bowl-shaped frontal depression and anterior margin of mesonotum with a row of large, forwardly-directed setae. Eyes situated very low on sides of head, their lower margin almost in contact with antennal socket. Upper margin of eyes well below level of the rounded vertex. Basal segment of abdomen with a transverse row of setae; long setae present laterally on either side of posterior end of abdomen.

Note. The very unusual shape of the head suggests that it is adapted for some biological activity unusual for a psocopteran. The circular outline of the head and the fact that it has an unusual arrangement of forwardly-directed setae (sensory?) suggests the possibility of phragmosis. This habit is confirmed for only one genus of wood-boring Psocoptera and suspected in another. Further observations on this species are needed to determine the function of the unusual head shape.

The larger of the two specimens (that from Salmon Beach) is in its final nymphal instar. This is indicated by two crescentic black marks on the front of the head in the position usually occupied by the ocellar tubercle in adults and the size of the wing buds, which extend to about half the length of the abdomen. The smaller specimen (from Stephens Reserve) is probably in the third instar, judging by the very short wing buds. The wing buds of the larger are brown. The sex of the larger specimen cannot be determined because the hind end of the abdomen is very membranous and pale and there is no obvious indication of genitalic structures. It is likely that the adults are macropterous (at least in one sex) and that the adult wings are coloured or patterned in some way. Full description and naming must await the collection of adults which, because of the unusual morphological features of the head, should be easily recognized as such, assuming that the adult resembles the nymphs in the unusual head shape described above.

**Distribution.** Known only from Lord Howe Island.

**PHILOTARSIDAE**

**Aaroniella howensis Smithers & Thornton**


**Material studied.** 1♂, eastern slope, Malabar Ridge, above Neds Beach, beating, 25.xi.2000, L. Wilkie, H. Smith.

**Distribution.** Known only from Lord Howe Island. The only other known specimen is a single female, the first psocopteran collected on the Island.

**Haplophallus tandus Smithers & Thornton**


**Distribution.** Known only from Lord Howe Island.
ELIPSOCIDAE

_Pentaclodus marmoratus_ Smithers & Thornton


Distribution. Known only from Lord Howe Island.

_Prionotodrilus parvus_ (Smithers & Thornton)


Distribution. Known only from Lord Howe Island.
MYOPSISOCIDAE

Nimbopsocus huttoni n.sp.

Material studied. ♀ Holotype (K232524), 4 ♀ ♀ (not designated paratypes as they are in poor condition), 8 nymphs, on rocks, northern Little Slope, 30.xi.2000, I. Hutton.

Etymology. This species is named for Dr Ian Hutton in recognition of his contribution to knowledge of the natural history of Lord Howe Island.

Description

Female. Coloration (in alcohol). Head pale brown with pattern of well defined, irregularly-shaped, dark, brown spots. Median epicranial suture pale, flanked on vertex by two rows of spots on each side. Four or five curved rows of spots on each epicranial plate, running almost parallel with inner margin of compound eye. Position of anterior arms of epicranial suture indicated by brown line (anterior arms themselves evanescent). Small brown spot below lower margin of eye behind which is a much larger spot which extends to occupy much of the gena near antenna base. Frons with a dark brown circle. Postclypeus with seven anteriorly converging brown stripes on each side of midline. Anteclypeus and labrum pale except for two small but obvious dark spots on anterior margin of latter. Scape, pedicel and first flagellar segment of antenna pale (antennae incomplete on all specimens). Eyes black. Ocelli black, integument between posterior ocelli pale. Maxillary palps pale except for light brown fourth segment. Mesothoracic antedorsum brown with pale median line; dorsal lobes pale with brown, ovoid area occupying middle part of lobe, median line very dark brown. Mesepisternum mostly dark brown, mesepimeron dark in dorsal part, pale in ventral part. Mesothoracic pleuron mostly pale except for darker area above coxa. Prothoracic legs with pale coxa, femur dark dorsally, irregularly marked laterally; tibia pale, basally and distally brown. Basal tarsal segment pale; second and third segments brown; claws very dark brown. Meso- and metathoracic legs similar to prothoracic legs but coxae laterally dark in basal half. Fore wing (Fig. 19) membrane mostly finely speckled with brown spots; main veins in basal half of wing, other than Cu2, with alternating lengths of dark and pale sections. Pattern on membrane is of denser spotting near marginal parts of cells R5, M1, M2, and M3. Hind wing membrane very faintly tinged with brown. Small spots at end of veins R1, R2+3; some sections of wing margin between end of R2+3 and M slightly darker than rest of margin. Abdomen ventrally pale, dorsally with strongly developed pattern of irregular, segmentally arranged, very dark brown spots of various sizes.

Morphology. Length of body not measured as all specimens have shrunken abdomens. Median epicranial suture distinct; anterior arms evanescent. Labrum with 8 external proximal sensilla and 7 marginal sensilla. Marginal sensilla consist of 4 trichoid and 3 placoid sensilla. Eyes small, not reaching level of vertex when seen from side. IO/D: 2.46; PO: 0.61. Ocelli small, anterior ocellus not obvious. Apex of lacinia with smaller inner tooth and bigger, broad outer tooth, apex of which is equipped with several small trichoid and placoid sensilla. Anterior arms of epicranial suture indicated by brown line (anterior arms themselves evanescent). Head pale brown with pattern of well defined, irregularly-shaped, dark, brown spots. Median epicranial suture pale, flanked on vertex by two rows of spots on each side. Four or five curved rows of spots on each epicranial plate, running almost parallel with inner margin of compound eye. Position of anterior arms of epicranial suture indicated by brown line (anterior arms themselves evanescent). Small brown spot below lower margin of eye behind which is a much larger spot which extends to occupy much of the gena near antenna base. Frons with a dark brown circle. Postclypeus with seven anteriorly converging brown stripes on each side of midline. Anteclypeus and labrum pale except for two small but obvious dark spots on anterior margin of latter. Scape, pedicel and first flagellar segment of antenna pale (antennae incomplete on all specimens). Eyes black. Ocelli black, integument between posterior ocelli pale. Maxillary palps pale except for light brown fourth segment. Mesothoracic antedorsum brown with pale median line; dorsal lobes pale with brown, ovoid area occupying middle part of lobe, median line very dark brown. Mesepisternum mostly dark brown, mesepimeron dark in dorsal part, pale in ventral part. Mesothoracic pleuron mostly pale except for darker area above coxa. Prothoracic legs with pale coxa, femur dark dorsally, irregularly marked laterally; tibia pale, basally and distally brown. Basal tarsal segment pale; second and third segments brown; claws very dark brown. Meso- and metathoracic legs similar to prothoracic legs but coxae laterally dark in basal half. Fore wing (Fig. 19) membrane mostly finely speckled with brown spots; main veins in basal half of wing, other than Cu2, with alternating lengths of dark and pale sections. Pattern on membrane is of denser spotting near marginal parts of cells R5, M1, M2, and M3. Hind wing membrane very faintly tinged with brown. Small spots at end of veins R1, R2+3; some sections of wing margin between end of R2+3 and M slightly darker than rest of margin. Abdomen ventrally pale, dorsally with strongly developed pattern of irregular, segmentally arranged, very dark brown spots of various sizes.

Morphology. Length of body not measured as all specimens have shrunken abdomens. Median epicranial suture distinct; anterior arms evanescent. Labrum with 8 external proximal sensilla and 7 marginal sensilla. Marginal sensilla consist of 4 trichoid and 3 placoid sensilla. Eyes small, not reaching level of vertex when seen from side. IO/D: 2.46; PO: 0.61. Ocelli small, anterior ocellus not obvious. Apex of lacinia with smaller inner tooth and bigger, broad outer tooth, apex of which is equipped with several small trichoid and placoid sensilla.

Figs 19–24. Nimbopsocus huttoni n.sp. Female: (19) fore wing; (20) epiproct; (21) gonapophyses; (22) subgenital plate; (23) paraproct; (24) entrance to spermatheca.
terminal rounded protuberances. Measurements of hind leg: F: 1.17 mm.; T: 1.87 mm.; t1: 0.62 mm.; t2: 0.08 mm.; t3: 0.1 mm.; rt: 7.75:1:1.25; ct. 22, 1, 1. Fore wing length: 5.0 mm.; fore wing width: 1.7 mm. Fore wing (Fig. 19) with Sc present as small, inconspicuous vestige, ending in costal cell. Rs and M meet in a point or are fused for a short length. Hind wing with Rs distal to separation from M strongly curved towards hind margin of wing then curving forward to division into R2+3 and R4+5; R2+3 reaches wing margin at wing apex. Rs and M fused for a length. M very strongly sinuous. End of Cu1 strongly recurved near wing margin. Epiproct (Fig. 20) with transverse band of irregularly spaced setae running more or less parallel to curved hind margin and a group of sortier, finer setae near middle of hind margin. Paraproct (Fig. 23). Subgenital plate (Fig. 22, drawn from non-type specimen) with short median posterior lobe bearing 3 terminal setae. Posterior part of plate glabrous, anteriorly plate with fine, scattered setae. Gonapophyses (Fig. 21) with ventral valve short, tapering distally to very fine point. Dorsal valve long, tapering, curved near distal end. Ninth sternite with simple sclerite at entrance to spermatheca (Fig. 24).

**Male. Unknown.**

**Nymphs.** Large nymphs are easily recognisable by their head pattern, which is similar to that of adults, the large, characteristic dark spot on the gena below the eye being obvious also in the nymphs. Knobbled glandular setae are present on abdomen and head.

**Distribution.** Known only from Lord Howe Island.

**Discussion**

Despite the lack of information on the male of *N. huttoni* it is placed in the genus *Nimbopsocus* by virtue of the close similarity of the female genitalia to those of the other species of the genus, especially the form of the sclerifications of the 9th abdominal sternite at the entrance to the spermatheca. The overall mottled wing pattern of *N. huttoni* is made up of much more evenly sized and spaced dark patches than any of the other species in the genus. The black genal patch is large and conspicuous in this species. It has a wing length similar to that of *N. australis* and *N. thorntoni* (female wing length: 5.0 mm) which both have longer wings than the smallest species of the genus, *N. hickmani* (female wing length: 3.4–3.6 mm).

**Nimbopsocus australis** (Brauer)


**Material studied.** 1 ♂, Stevens Reserve, nr. Signal Point, yellow pan trap, 8–12.xii.1988, D.C.F. Rentz. Specimen in Australian National Insect Collection.

**Distribution.** First record for Lord Howe Island. Previously known from Australia, Tasmania, Norfolk Island, New Zealand, Solomon Islands, Kermadecs, ?India. The presence of this species in India requires confirmation. Found mainly on bark, cut timber and paling fences carrying algal and fungal growth.
ACKNOWLEDGMENTS. I would like to thank Professor E.L. Mockford for helpful discussion of the spermathecal structures of *Lepolepis* species and on the family placing of *Trogium nigrum*, those responsible for collecting and sorting the many specimens, Elizabeth Jefferys for pointing out discrepancies in botanical nomenclature, Lance Wilkie for providing electronically handled collection data and Gerry Cassis for making the Lord Howe Island collections available for study.

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